

# SAMPLE SUMMARY

Lambda DNA (<https://www.thermofisher.com/order/catalog/product/SD0021?SID=srch-srp-SD0021>)




was incubated with methyltransferases:

M.TaqI (TCGA<sub>A</sub>, N6-A)

M2.BcnI (CCSGG, N4-C)

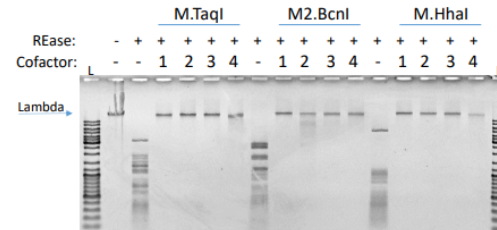
M.HhaI (GC<sub>C</sub>GC, 5-C) (engineered variant)

and cofactors, each MTase with:

- |                            | Transferable group  |
|----------------------------|---|
| 1. AdoMet                  | -CH <sub>3</sub>  |
| 2. AdoButyn                |  |
| 3. AdoHexynN <sub>3</sub>  |  |
| 4. AdoHexynNH <sub>2</sub> |  |

The reaction mixtures were then treated with Proteinase K and column-purified (Zymo Research, genomic DCC), eluted with 100 µl Elution Buffer (10 mM Tris-HCl, pH 8.5, 0.1 mM EDTA ).

The extent of DNA modification was assayed by protection from the cognate restriction endonuclease (i.e. R.TaqI, R.BcnI and R.Hin6I, respectively)



Samples with complete protection (nearly all the target sites have an attached group): M.TaqI with cofactors 1, 2 and 3; M2.BcnI – 1, 3 and 4; M.HhaI 1, 2 and 3.

The rest (TaqI 4, BcnI 2 and HhaI 4) have incomplete protection, i.e. not all the recognition sites are modified.

# SEQUENCING SUMMARY

Note: we can only sequence one sample at a time at the moment, which we do as spike-ins into other runs; we have now obtained the ONT barcoding kit so we hope to be able to run all remaining samples soon in a single run

Species	Assay	#	Mapping	Base Contexts	Reads	Total Bases	Mean Read Length	Median Read Length
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## TaqI + HexynNH<sub>2</sub>

Lambda_NEB	alkylated DNA	2018_12_13_0h_Diamide_Lithu_TaqI_Hey-Lambda	Tombo 1.3 denovo	CG-only	31,857	139,682,998	4,385	2,998
Lambda_NEB	alkylated DNA	2018_12_13_0h_Diamide_Lithu_TaqI_Hey-Lambda	Tombo 1.3 denovo	GC-only	31,857	139,971,785	4,394	3,002
Lambda_NEB	alkylated DNA	2018_12_13_0h_Diamide_Lithu_TaqI_Hey-Lambda	Tombo 1.3 denovo	T-only	31,857	140,686,793	4,416	3,023
Lambda_NEB	alkylated DNA	2018_12_13_0h_Diamide_Lithu_TaqI_Hey-Lambda	Tombo 1.3 denovo	all	31,857	140,816,187	4,420	3,027
Lambda_NEB	alkylated DNA	2018_12_13_0h_Diamide_Lithu_TaqI_Hey-Lambda	Tombo 1.3 denovo	m6A-GC-CG-only	31,857	140,781,227	4,419	3,026
Lambda_NEB	alkylated DNA	2018_12_13_0h_Diamide_Lithu_TaqI_Hey-Lambda	Tombo 1.3 denovo	m6A-only	31,857	140,683,923	4,416	3,022

## TaqI + CH<sub>3</sub>

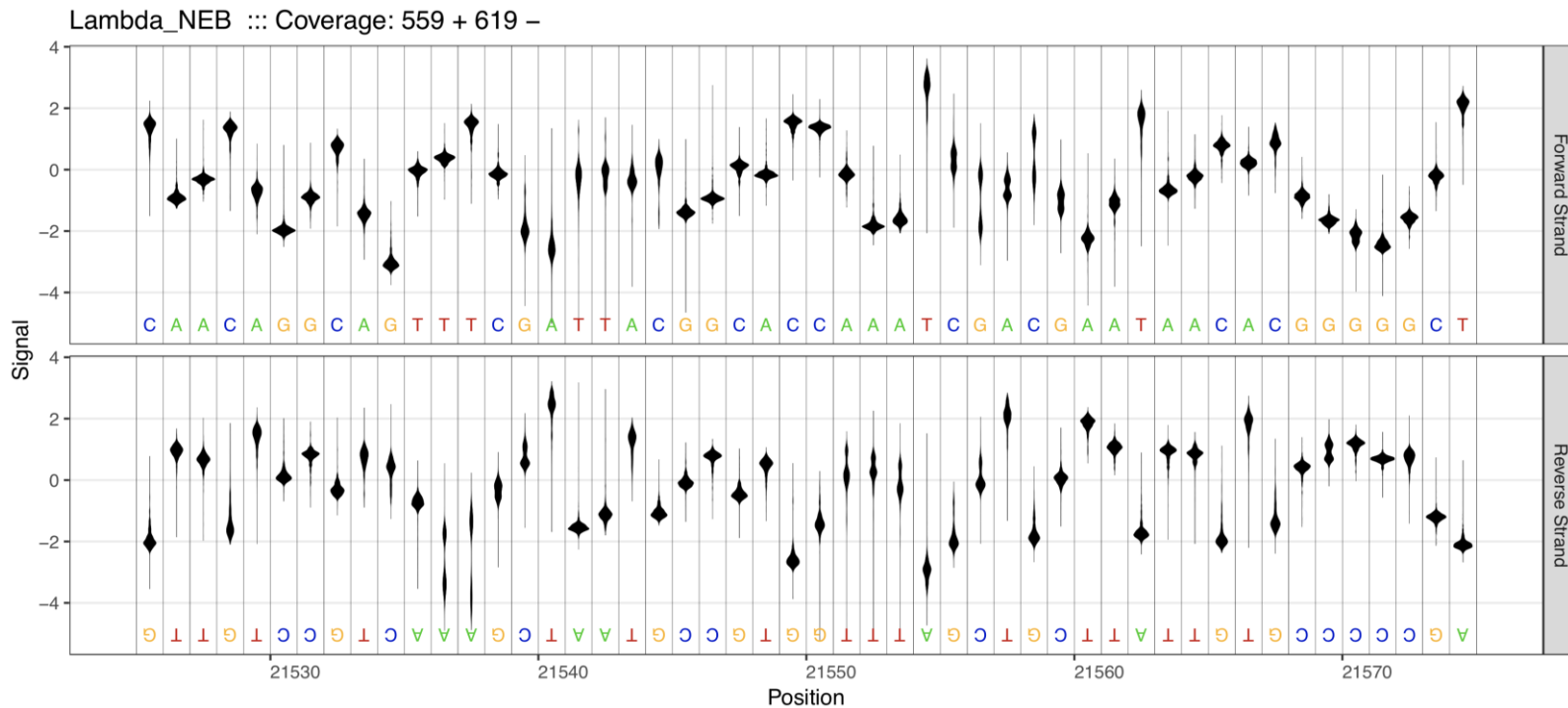
Lambda_NEB	alkylated DNA	2019_01_16_60min_Diamide_Lithu_TaqI_Methyl	Tombo 1.3 denovo	CG-only	2,909	16,978,041	5,836	3,861
Lambda_NEB	alkylated DNA	2019_01_16_60min_Diamide_Lithu_TaqI_Methyl	Tombo 1.3 denovo	GC-only	2,909	17,000,315	5,844	3,865
Lambda_NEB	alkylated DNA	2019_01_16_60min_Diamide_Lithu_TaqI_Methyl	Tombo 1.3 denovo	T-only	2,909	17,064,980	5,866	3,897
Lambda_NEB	alkylated DNA	2019_01_16_60min_Diamide_Lithu_TaqI_Methyl	Tombo 1.3 denovo	all	2,909	17,076,734	5,870	3,901
Lambda_NEB	alkylated DNA	2019_01_16_60min_Diamide_Lithu_TaqI_Methyl	Tombo 1.3 denovo	m6A-only	2,909	17,064,555	5,866	3,894

## TaqI + Butyn

Lambda_NEB	alkylated DNA	2019_01_29_Lambda_Butyl	Tombo 1.3 denovo	CG-only	26,414	82,919,697	3,139	3,412
Lambda_NEB	alkylated DNA	2019_01_29_Lambda_Butyl	Tombo 1.3 denovo	GC-only	26,414	82,170,164	3,111	3,375
Lambda_NEB	alkylated DNA	2019_01_29_Lambda_Butyl	Tombo 1.3 denovo	T-only	26,414	83,287,319	3,153	3,421
Lambda_NEB	alkylated DNA	2019_01_29_Lambda_Butyl	Tombo 1.3 denovo	all	26,414	83,440,725	3,159	3,425
Lambda_NEB	alkylated DNA	2019_01_29_Lambda_Butyl	Tombo 1.3 denovo	m6A-only	26,414	83,336,353	3,155	3,423

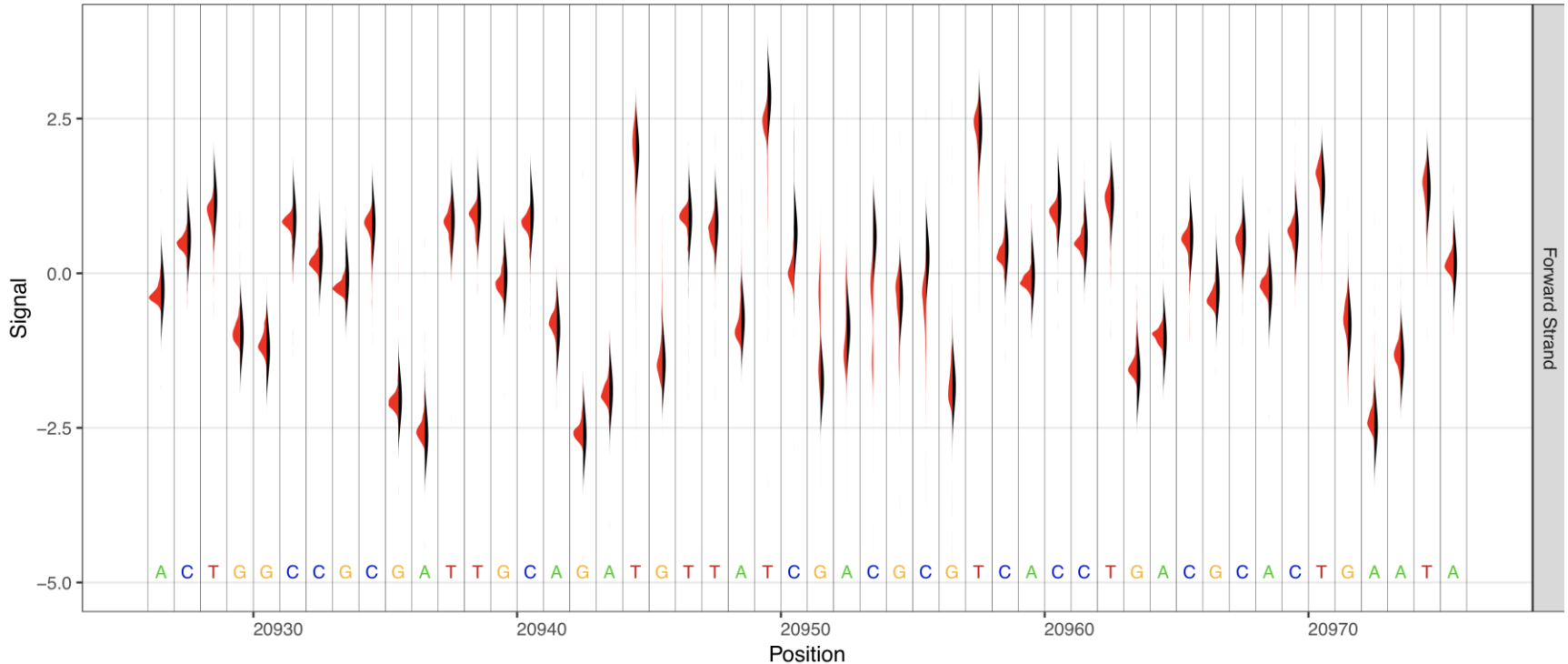
# TAQI HexynNH<sub>2</sub> RESULTS

- High coverage (559X on the Plus strand and 619X on the Minus strand) with two TCGA sites
- Shown are the distributions of the Nanopore signal for each base. Most bases have a very tight distribution, the 2-3 bases around the A of TCGA have a bimodal distribution.
- We don't know if the bimodal distribution comes from less than 100% enzymatic efficiency of TaqI or bad base calling

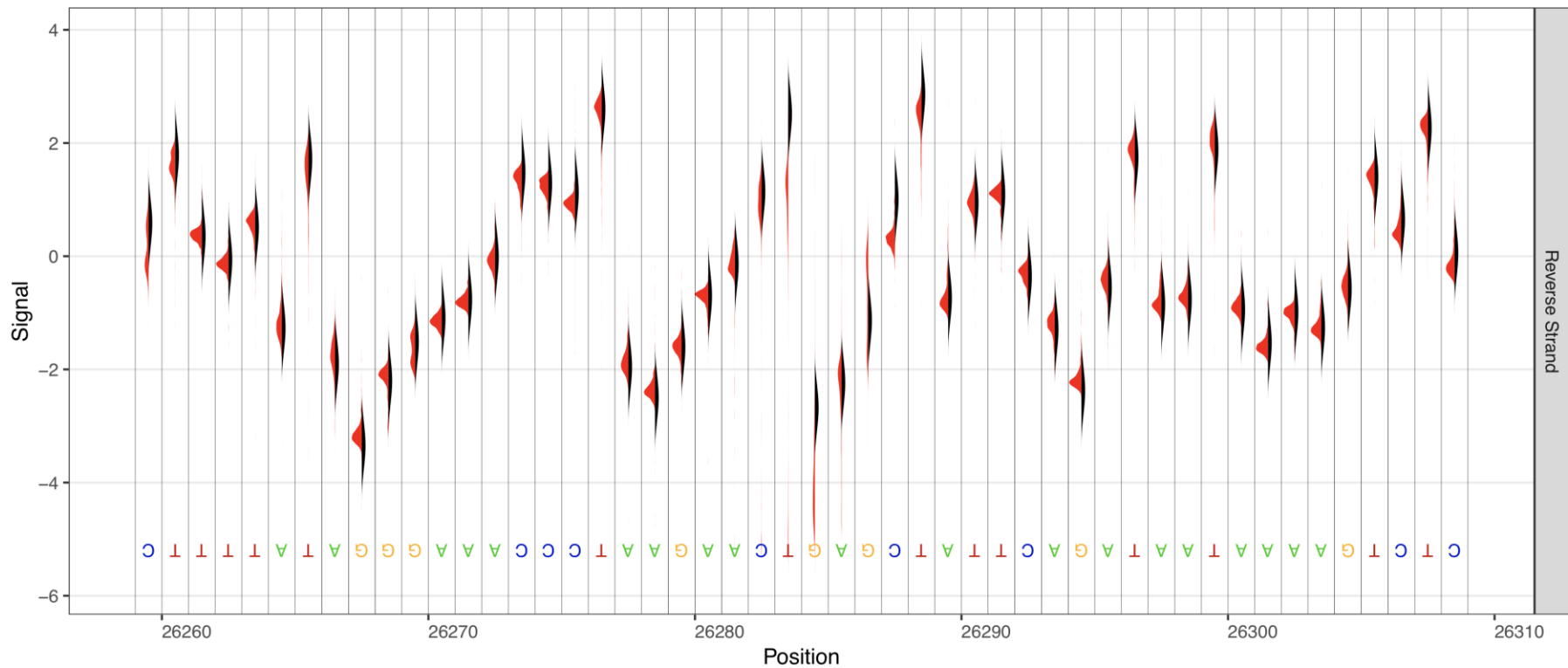


- Here we are looking at the distribution of signal for a high coverage region that has a TCGA motif in it. But now we compare the distribution to the model (the more off the model, the more modified the base is called)
- The modified A causes a big shift in the signal for the bases around it

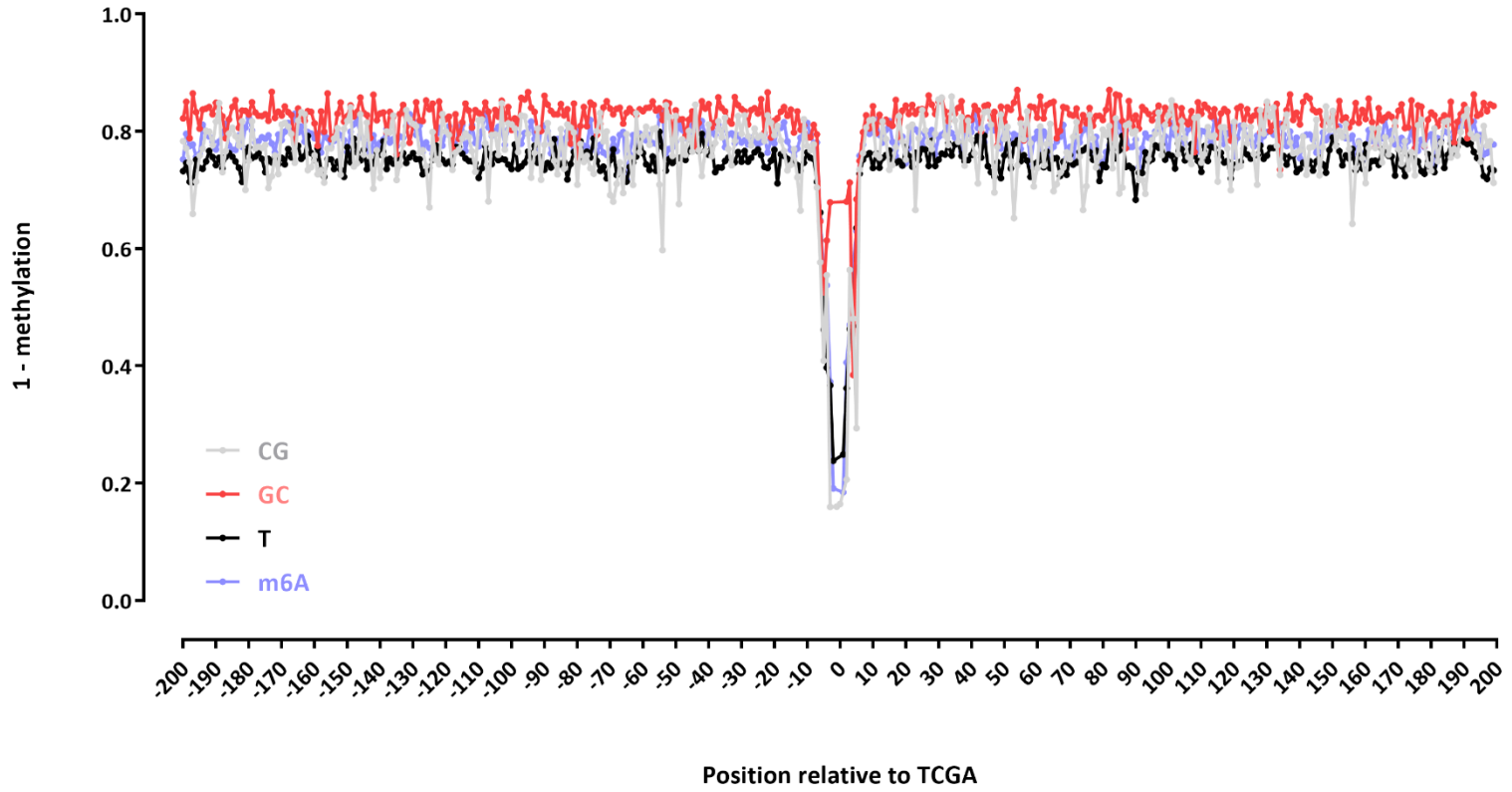
Lambda\_NEB:+ Est. Frac. Alternate: 0.99 Coverage: 163 ::: Coverage: 556 (Model in Black)



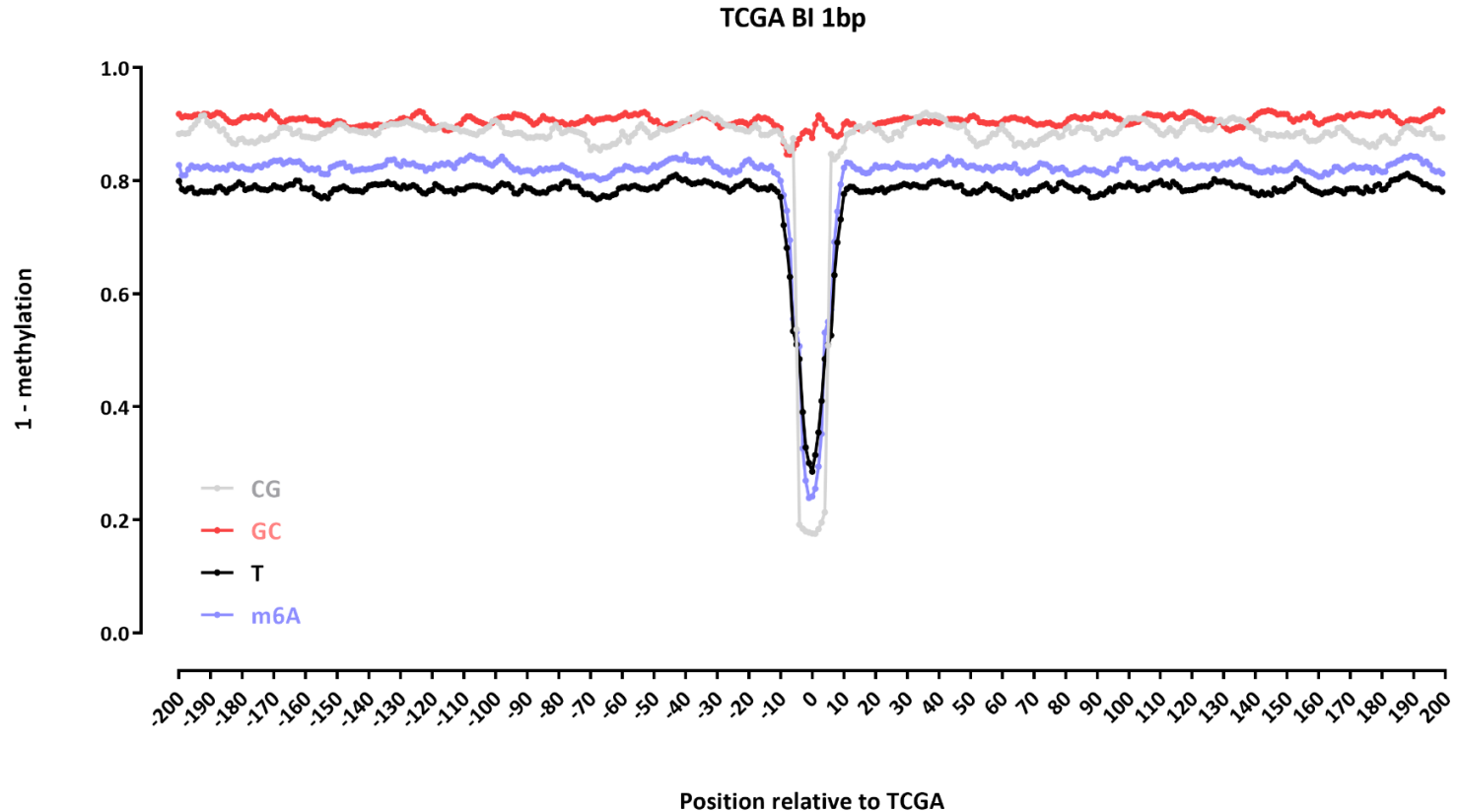
Lambda\_NEB:- Est. Frac. Alternate: 0.98 Coverage: 224 ::: Coverage: 668 (Model in Black)



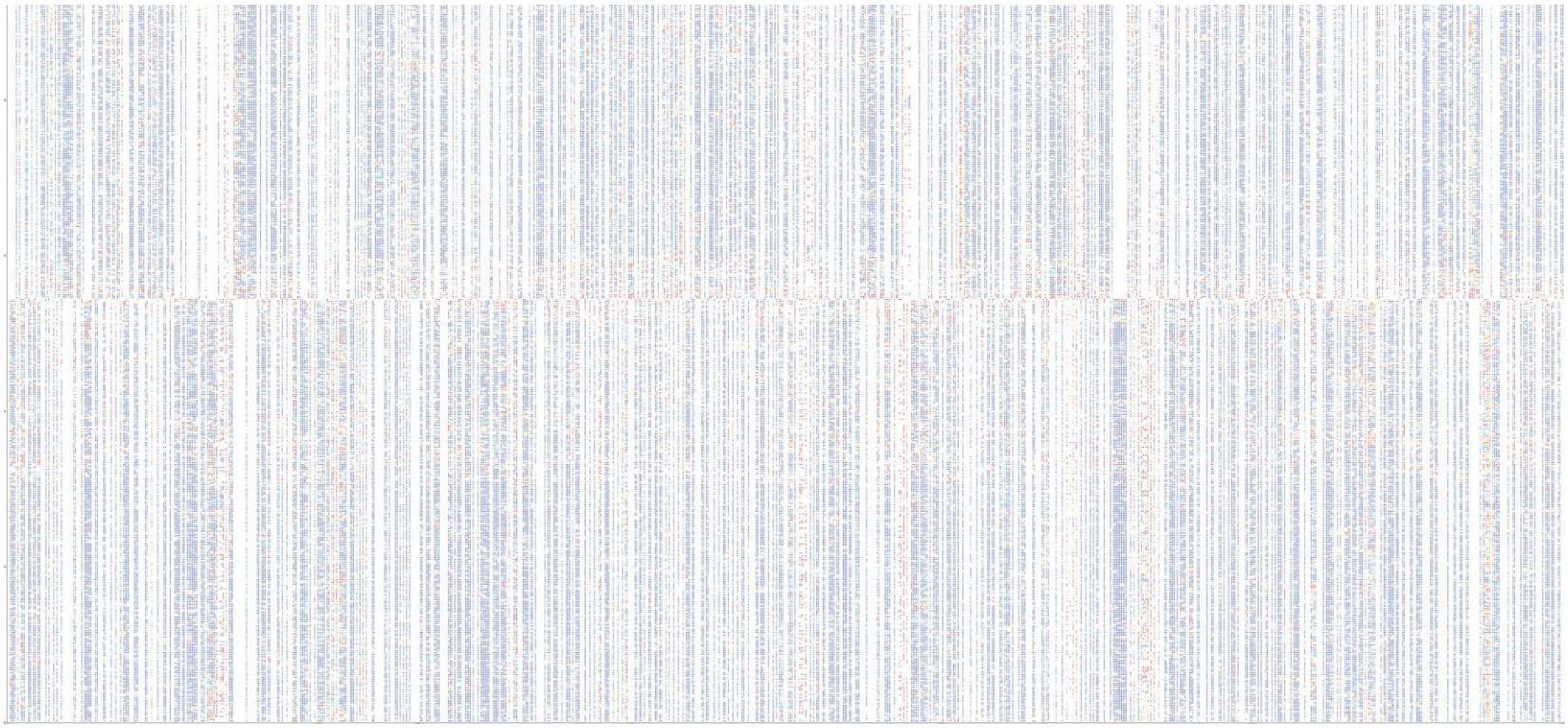
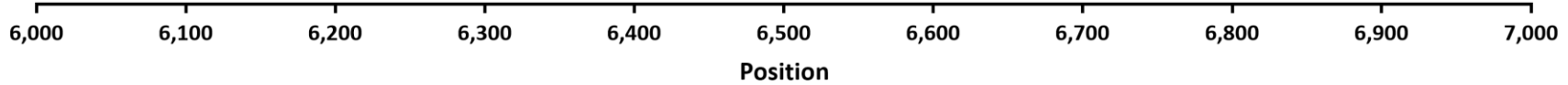
### TCGA 1bp



- “BI” means “Bayesian integration”; we aggregate signal across neighboring bases using a Bayesian procedure to obtain more reliable calls of accessibility (in the context of chromatin). This is done over 10bp windows for every base in the sequence



1000 reads



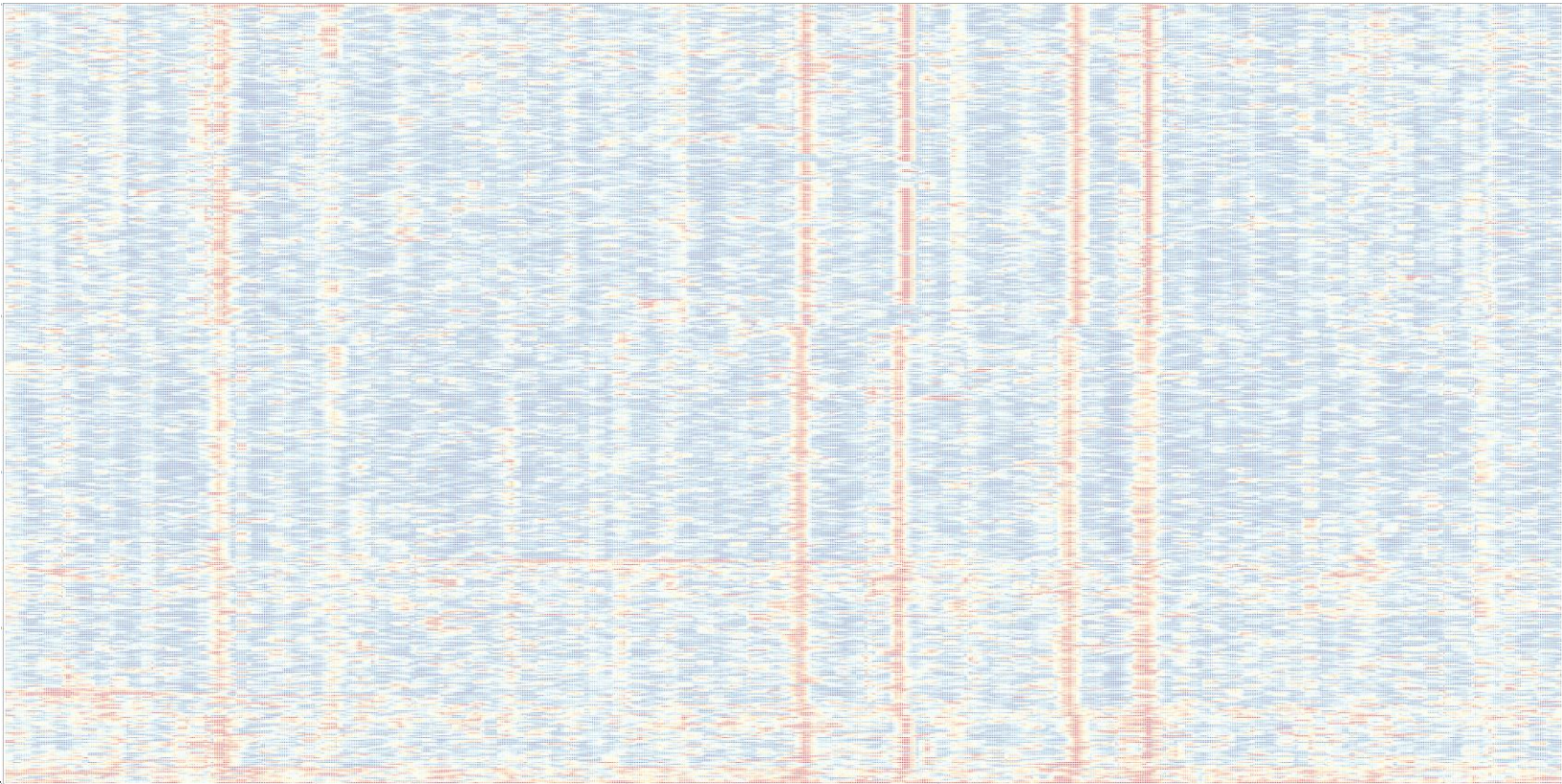
**BI**



6,000 6,100 6,200 6,300 6,400 6,500 6,600 6,700 6,800 6,900 7,000

Position

1000 reads

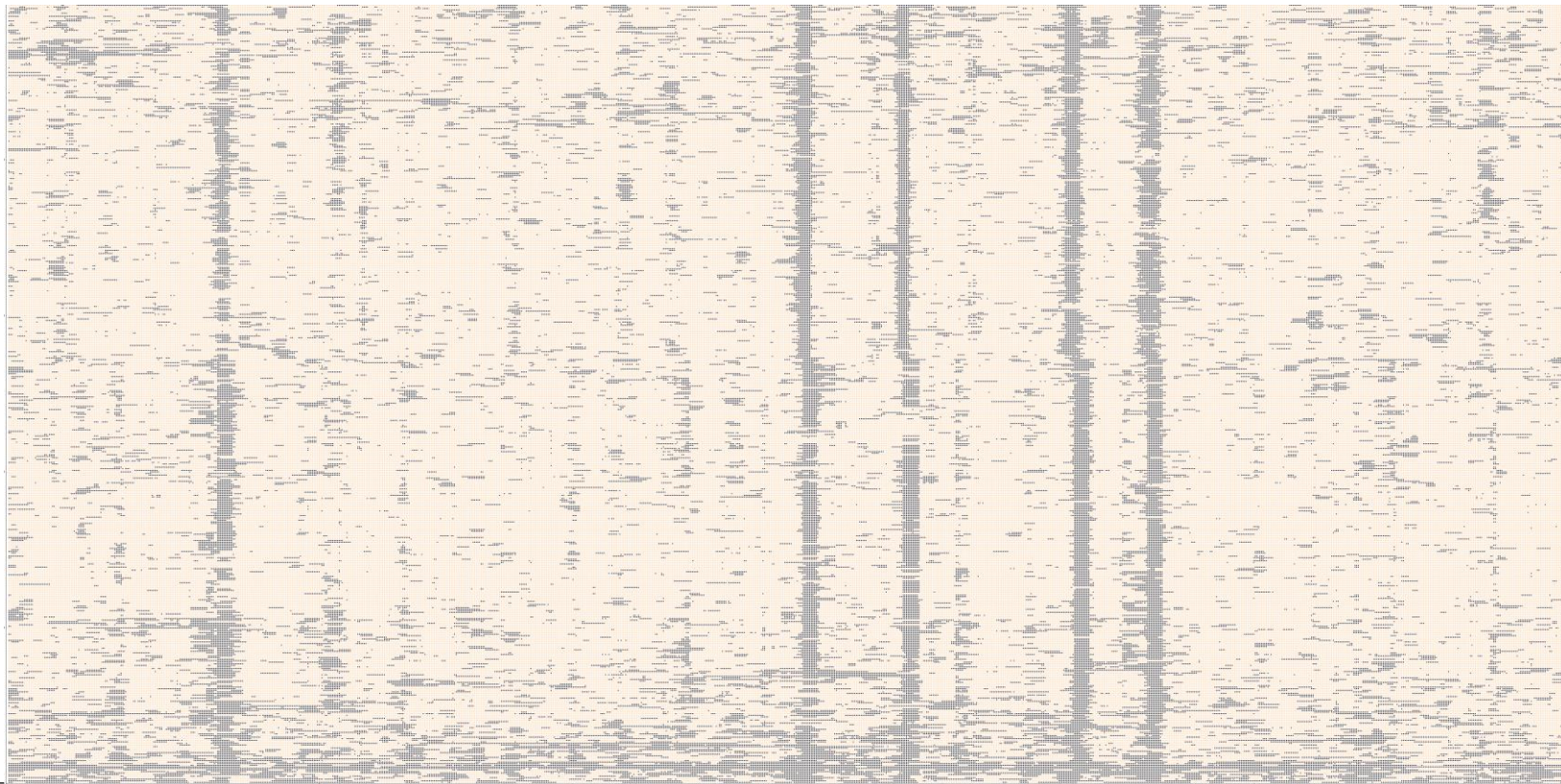


9

**BI**  
**P < 0.5**

6,000 6,100 6,200 6,300 6,400 6,500 6,600 6,700 6,800 6,900 7,000

Position



**BI**  
**P < 0.3**

|

|

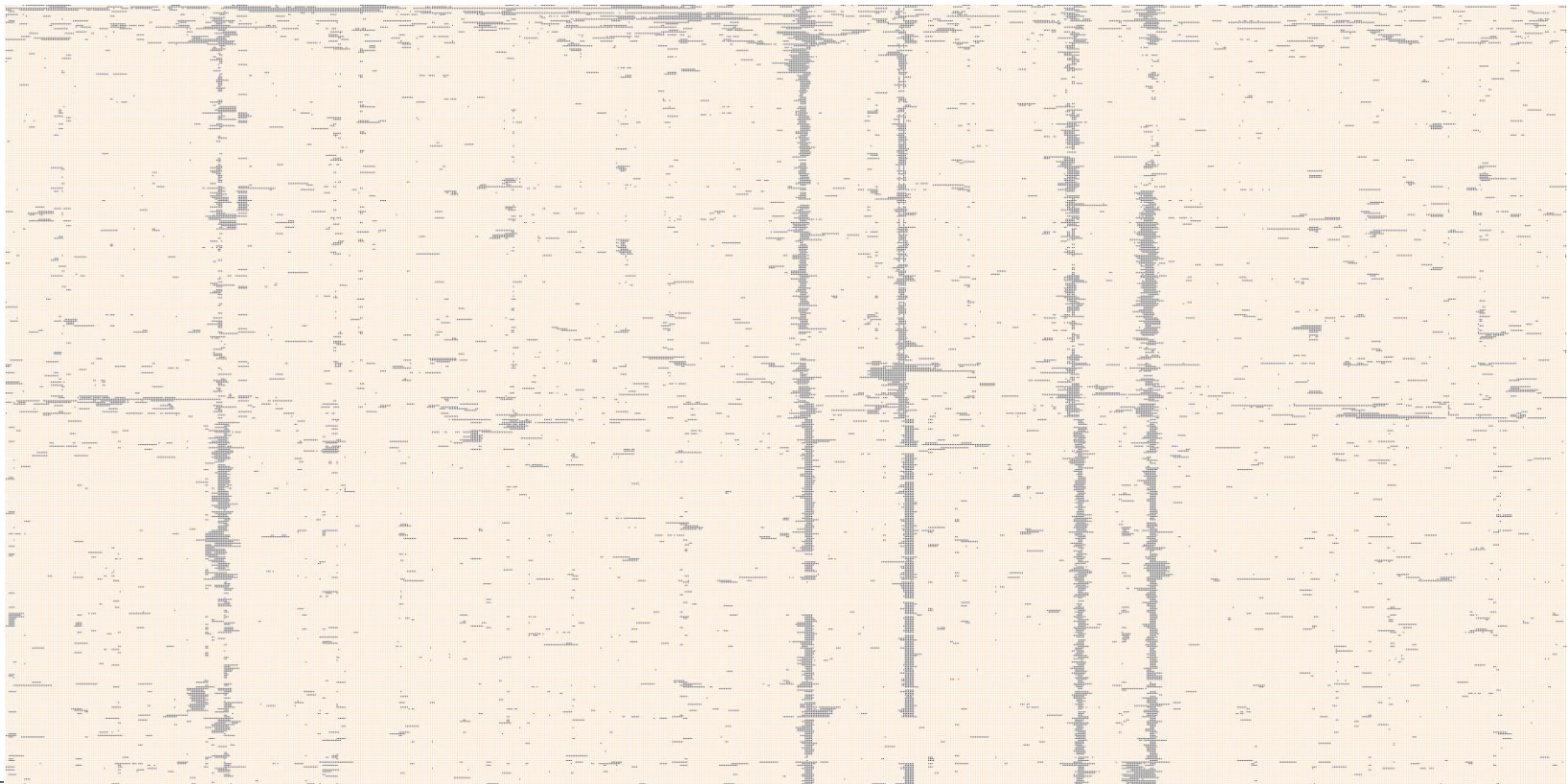
|

|

|

6,000 6,100 6,200 6,300 6,400 6,500 6,600 6,700 6,800 6,900 7,000

Position



**BI**  
**P<0.1**

|

|

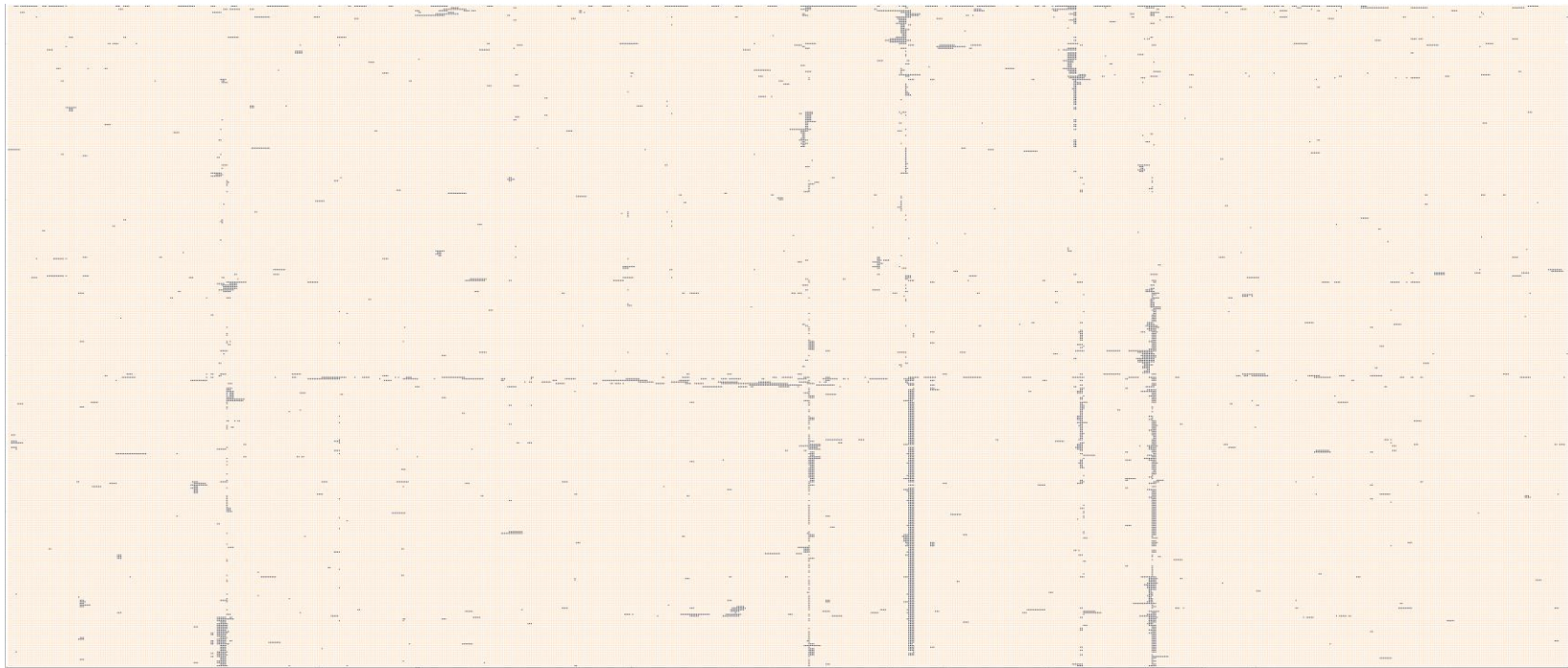
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|

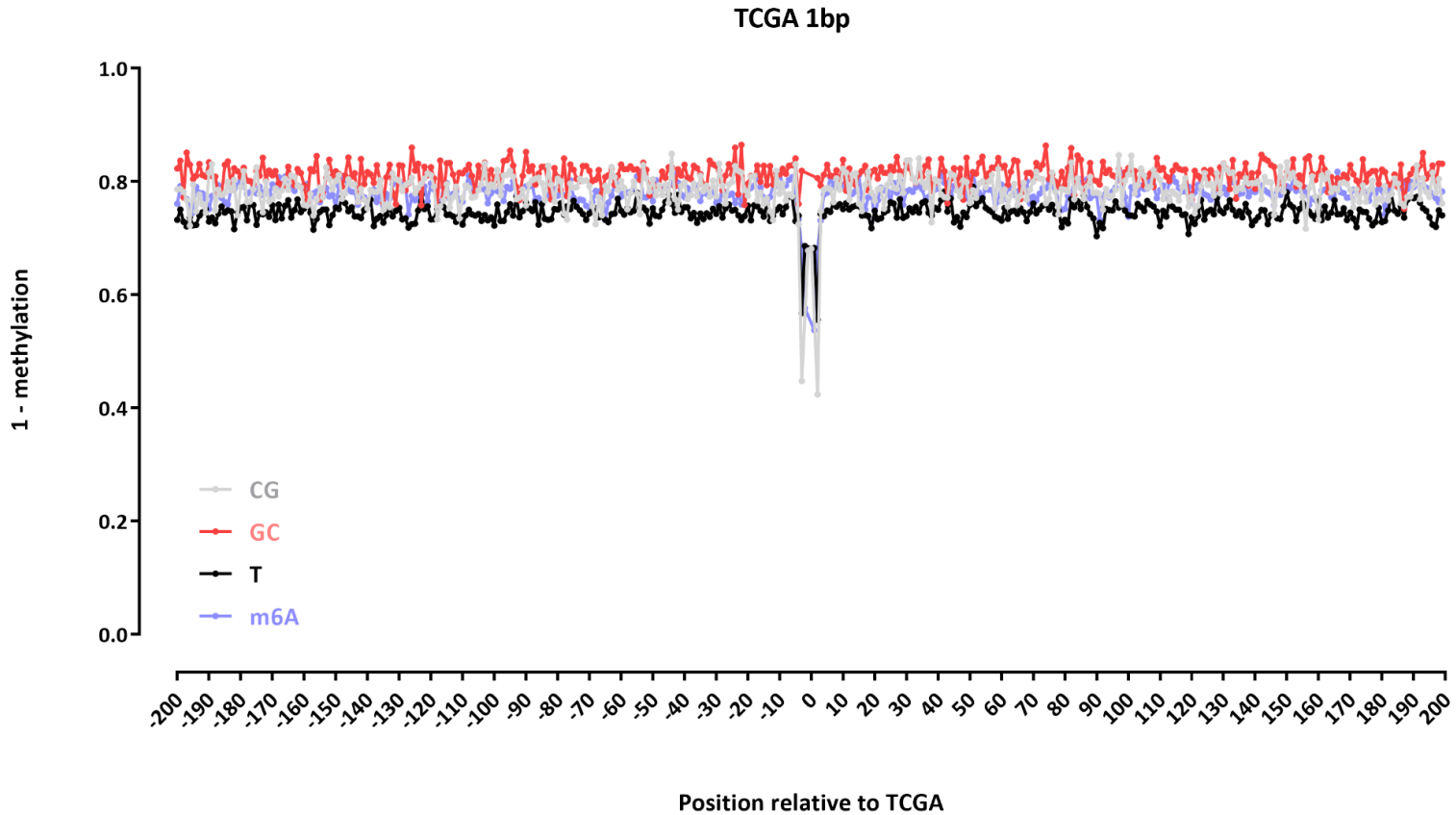
|

6,000 6,100 6,200 6,300 6,400 6,500 6,600 6,700 6,800 6,900 7,000

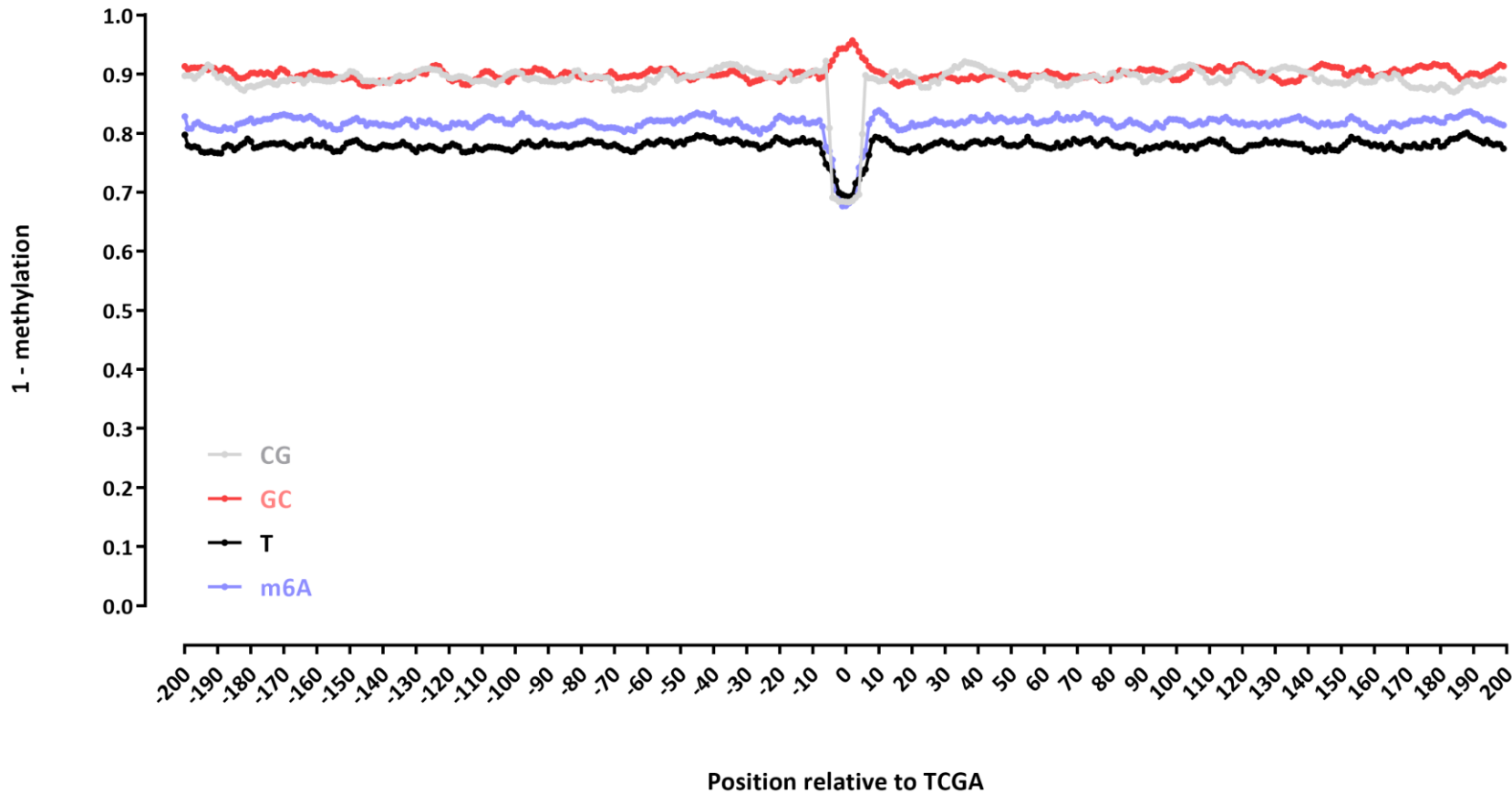
Position

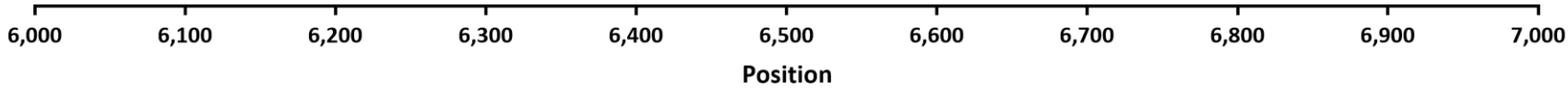


# TAQI CH<sub>3</sub> RESULTS

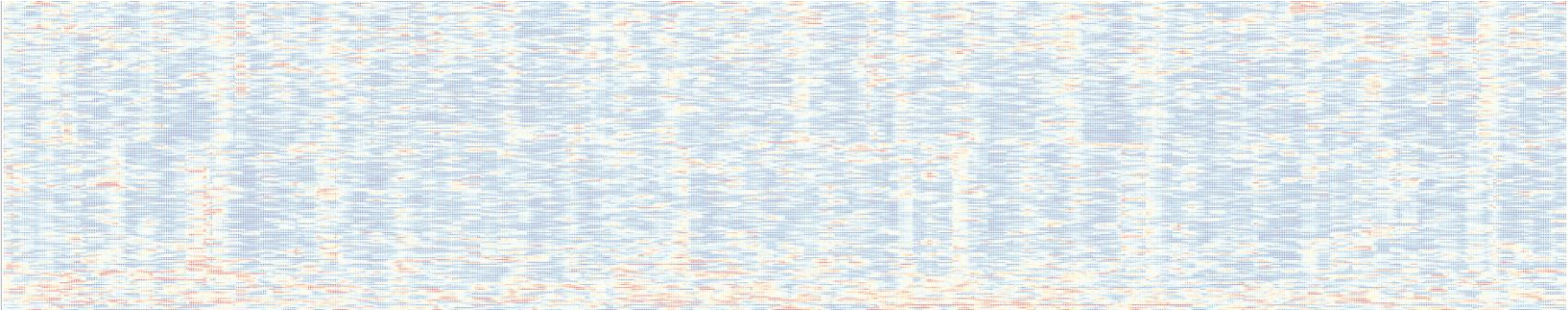


### TCGA BI 1bp





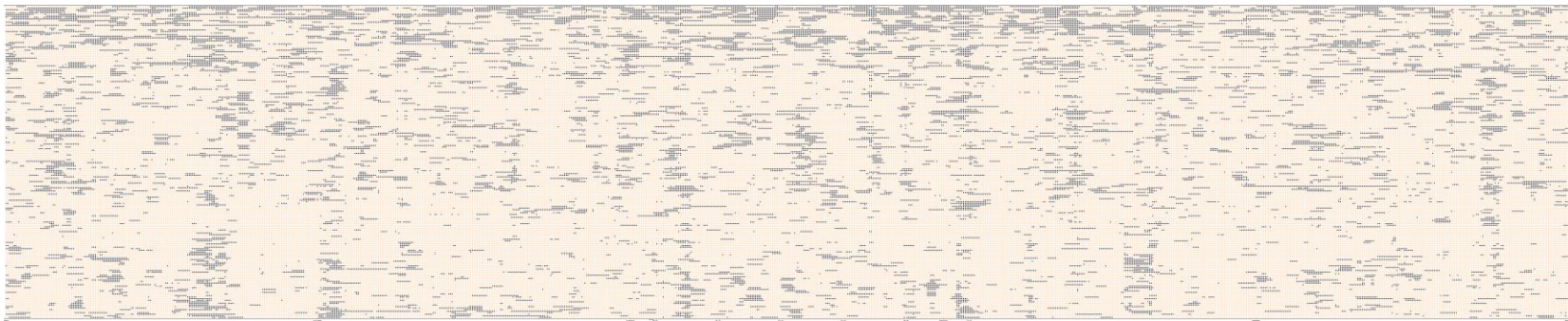
200 reads



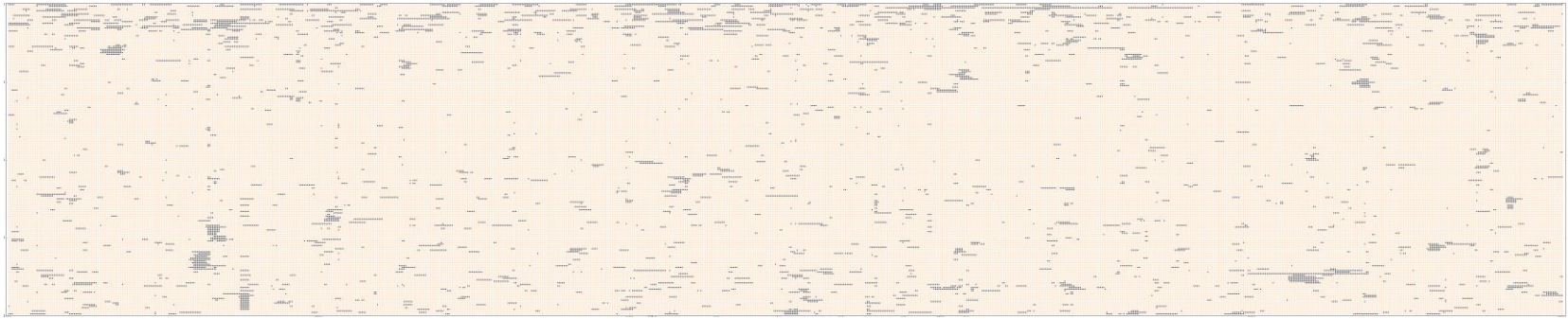
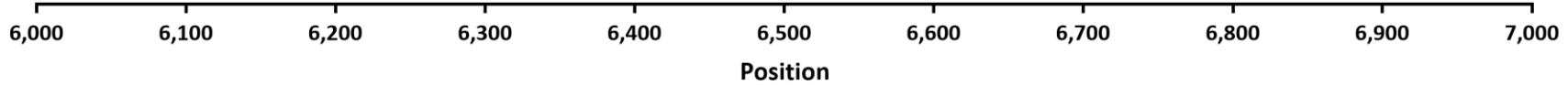
**BI**  
**P < 0.5**

6,000 6,100 6,200 6,300 6,400 6,500 6,600 6,700 6,800 6,900 7,000

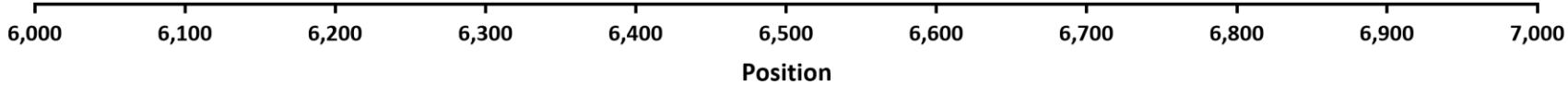
Position



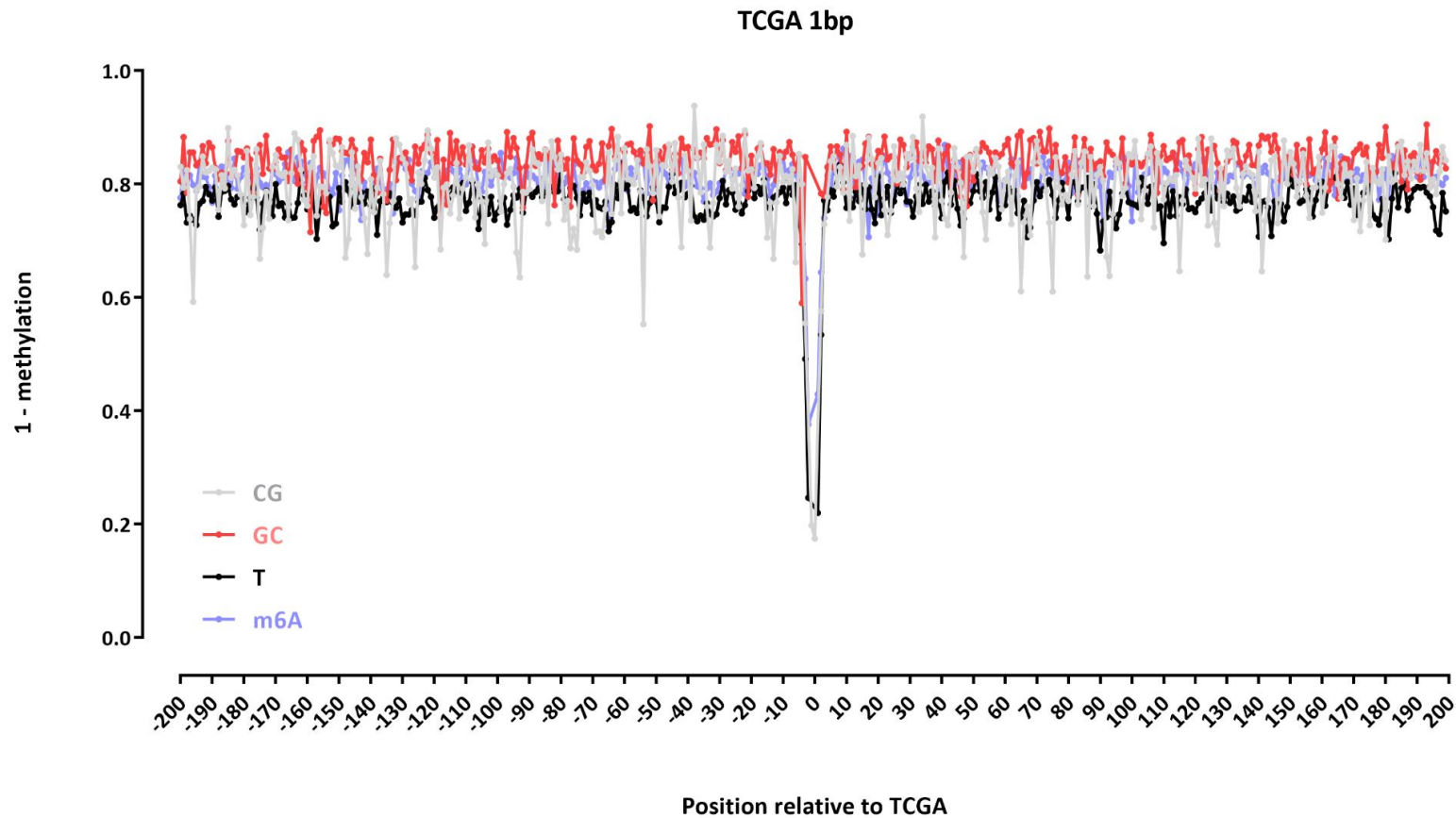
**BI**  
**P<0.3**



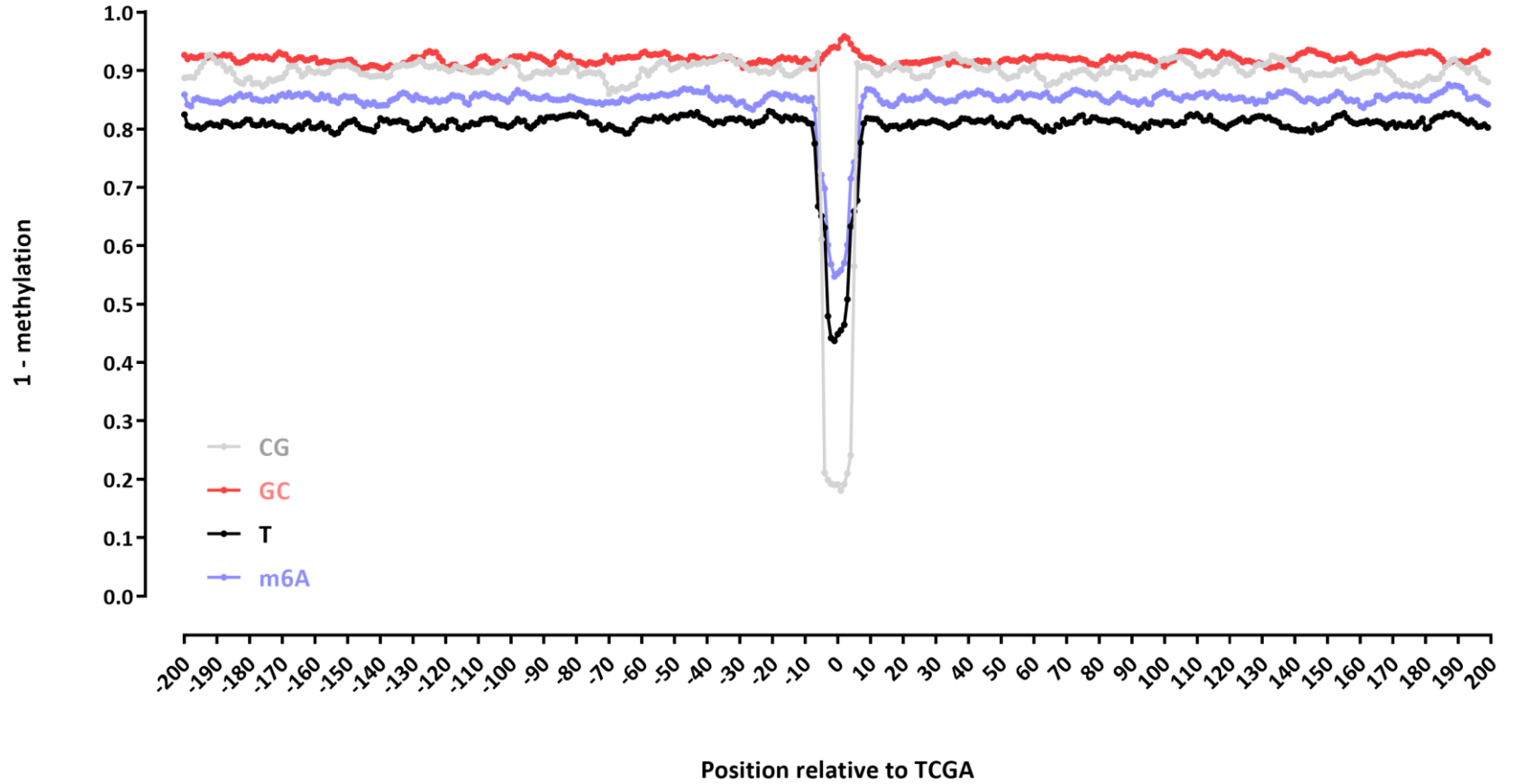
**BI**  
**P<0.1**



# TAQI Butyn RESULTS



# TCGA BI 1bp



**BI**

|

|

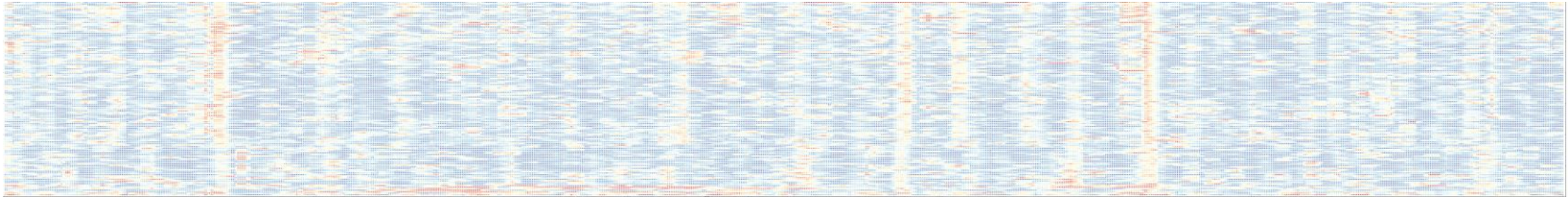
|

|

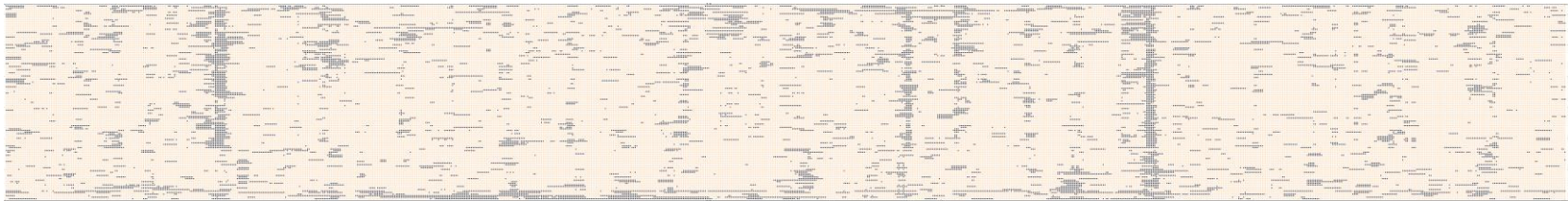
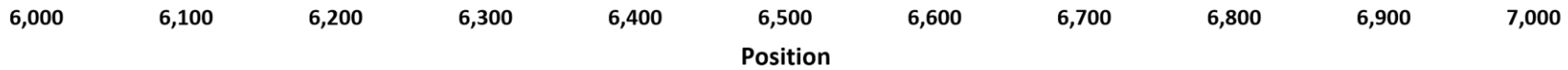
|

6,000 6,100 6,200 6,300 6,400 6,500 6,600 6,700 6,800 6,900 7,000

Position



**BI**  
**P<0.5**



**BI**  
**P<0.3**

